NOTA





www.ajaes.ufra.edu.br



AUTORES:

Iracema Nunes de Barros¹ Nairléia dos Santos Silva¹

Maria das Graças Avila Ribeiro Almeida²

Antonio Vicente Magnavita Anunciação²

Sonia da Silva Laborda²

Elizabeth Juvêncio Ramalho²

Eugênia Márcia de Deus Oliveira²

¹USP, FMVZ, Av. Prof. Orlando Marques de Paiva, 87, 05508-270, SP,SP Brasil

²UFBA, Av. Ademar de Barros, 500, 40170- 110, Salvador, Bahia, Brasil.

Recebido: 09/09/2009 Aprovado: 07/06/2010

AUTOR CORRESPONDENTE:

Iracema Nunes de Barros E-mail: cemavet@gmail.com

PALAVRAS-CHAVE:

Lentivírus, Visna/maedi, Pequenos ruminantes, Sorologia.

KEY WORDS:

Lentivirus, Visna/maedi, Small ruminants, Serology.

Detection of antibodies to Visna/Maedi in sheep from Recôncavo Baiano

Ocorrência de anticorpos contra Visna/Maedi em ovinos na região do Recôncavo Baiano

Abstract: The Visna/Maedi virus is a Lentivirus that causes a multisystem, progressive, slow-developing disease that can affect sheep and goats. After long periods of subclinical infection, the disease can slowly progress to cause the degeneration of various organs, cachexia and death. The diagnosis is made by the observation of clinical signs and confirmed by serological tests and isolation and/or identification of the virus. There is no treatment for the disease. The control is based on segregated breeding, handling and slaughtering of the positives. As Visna/ Maedi is on the list of the World Organization for Animal Health (OIE), reporting of the disease is obligatory and it has been found in some States in Brazil, causing negative economic impacts with its spread. Given the limited seroepidemiological data available in the Brazilian State of Bahia, this study aimed to carry out a seroepidemiological survey to investigate the occurrence of anti-Visna/Maedi virus antibodies in sheep from Recôncavo Baiano. Two hundred serum samples from sheep were subjected to the agar gel immunodiffusion test (AGID). None of the serum samples were found to be positive. To confirm the absence of Visna/Maedi in sheep herds in Bahia, there is a need for further studies with significant sampling including a more sensitive assay.

Resumo: O Visna/Maedi virus (MVV) é um Lentivirus que causa uma enfermidade multissistêmica, progressiva e de curso lento, que pode acometer ovinos e caprinos. Após longos períodos de infecção assintomática, pode progredir lentamente à degeneração de vários órgãos, caquexia e morte. O diagnóstico se faz através da observação de sinais clínicos e a confirmação mediante testes sorológicos, isolamento e/ou identificação do vírus. Não há tratamento, até o momento. O controle é baseado na criação segregada, manejo e sacrifício dos positivos. Por fazer parte da lista da Organização Mundial de Saúde Animal (OIE), ser uma doença de notificação compulsória, ter sido verificada em alguns estados do Brasil, podendo causar impactos econômicos negativos com a sua disseminação, e haver escassos dados soroepidemiológicos no Estado da Bahia, esta pesquisa teve por objetivo realizar inquérito sorológico para investigar a ocorrência de anticorpos para Visna/Maedi em ovinos, na região do Recôncavo Baiano. Duzentas amostras de soros ovinos foram submetidas ao teste de imunodifusão em gel de agarose (Idag). Nenhum dos soros pesquisados foi reagente. Para que se possa realmente confirmar a ausência de Visna/Maedi nos rebanhos de ovinos na Bahia, há necessidade de outros estudos com amostragem significativa e a inclusão de testes mais sensíveis.

1 Introduction

Small Ruminant Lentivirus (SRLV) is a generic term used to describe the etiologic agents of caprine arthritis encephalitis (CAE) and Visna/Maedi (Visna). These viruses belong to the *Retroviridae* family and affect goats and sheep, are widespread and phenotypic, biological and antigenically related (PASICK, 1998). The Visna is a disease of sheep and it is chronic, progressive and multisystemic, which mainly affects lungs, mammary glands, brain and more rarely, joints (PUGH, 2002), causing economic losses due to the decrease in productive life and reproductive efficiency, delay in growth, increased mortality of offspring, and indirect losses resulting from the devaluation of the herds, early replacement of diseased animals, costs control measures and possible trade barriers, which may limit or prevent the international trade (SILVA et al., 2005).

The SRLV transmission is mainly via ingestion of colostrum and milk of infected females or through the respiratory contact, more common in intensive system or via contaminated semen though with lesser risk (BLACKLAWS et al., 2004). According to Leginagoikoa et al (2006), there is evidence that horizontal transmission is very low, despite the prolonged direct contact between infected and uninfected sheep, highlighting the need to investigate the dynamics of virus excretion and transmission among animals to obtain grants and to establish control in endemic populations of Visna. Furthermore, phylogenetic studies have demonstrated the SRLV transmission in sheep to goats and vice versa (LEROUX et al., 1997; SHAH et al., 2004; GJERSET et al., 2009; GLARIA et al., 2009).

Diagnosis of SRLV can be made through observation of clinical signs, but due to the SRLV characteristics of persistent infection and no symptoms in most infected animals, the confirmation is made through serologic tests, or isolation or identification of the virus. The most way extensively used to diagnose SLRV is to perform serology by the agar gel immunodiffusion (AGID), due to be recommended by the World Organisation for Animal Health (OIE, 2004), to present low cost and high specificity and practicality (PUGH, 2002). However, the test has low sensitivity in the early phase of infection (LEROUX et al., 1997; FLEET et al., 2005; ELTAHIR et al., 2006).

So far, there is no treatment for SRLV and control is based on the reduction of infection with the implementation of appropriate management with segregated breeding and slaughter of the positives (SMITH, 1994; REINA et al., 2009).

Several authors reported the presence of antibodies to Visna in Brazil and identified in Rio Grande do Sul (MOOJEN et al., 1995; RAVAZZOLO et al., 1995; RAVAZZOLO et al., 2001); in São Paulo (FERNANDES et al., 2003); in Rio Grande do Norte (SILVA et al., 2003); in Ceará (ALMEIDA et al., 2003; ARAÚJO et al., 2004); in Pernambuco (OLIVEIRA et al., 2006; COSTA et al., 2007). However, the first reports in the State of Bahia occurred recently in Juazeiro by Souza et al. (2007) and Martinez et al. (2008).

Thus, considering the negative economic impact caused by spread of Visna, and the disease is included in list of OIE as a disease of compulsory notification, and has already been verified in some States of Brazil, and the lack of seroepidemiologic data in Bahia, this research aimed to carry out a seroepidemiological survey to investigate the occurrence of antibodies to Visna/Maedi virus (Visna) in sheep from Recôncavo Baiano Region.

2 Material and Methods

Two hundred sheep sera samples from various breeds were examined, convenience sampling, included 43 males and 157 females, aged five months to eight years. There was no criterion of probability applied to establish the number of animals in this study. In this way, all analyzed samples were harvested from nine properties as belonging to eight municipalities of the Recôncavo Baiano Region: Santo Amaro, Castro Alves, Jaguaripe, Jequiriça, Muritiba, Governador Mangabeira, Mutuípe and Laje. These municipalities were contemplated according to the acceptance of farm owners and the ease of access; therefore, there was no criterion for selection of sampling. The collection of samples was carried out from October 2006 until January 2007. In some herds, animals with respiratory symptoms were identified coexisting with asymptomatic animals.

Blood samples were harvested in sterile glass tubes without anticoagulant by puncture of the external jugular vein, and a total volume of 10 mL per animal were obtained. The tubes were placed in vertical position to the coagulation, and identified with numbers and properties names, and then packed in styrofoam boxes filled with dry ice and sent to the Laboratories of Bacteriosis and Virosis of the Veterinary Medicine School/ UFBA. After clot

retraction, the blood samples were centrifuged at 2000 g for 10 minutes, and the sera were placed in 1.5 mL polypropylene microtubes (Eppendorf) at -20 °C until the performance of serological tests (ALMEIDA et al., 2003; OLIVEIRA et al., 2006). At the time of collection of the biological material, a clinical evaluation was carried out in the animals to detect suggestive alterations of Visna infection.

Serology for detection of antibodies against Visna was carried out by the AGID (OIE. 2004), using a kit produced by the Federal Rural University of Pernambuco, consisting of antigen produced in cell cultures infected with Visna (protein - p28), and positive serum harvested from naturally infected animal. The AGID test was performed according to the manufacturer's recommendations and followed the OIE recommended pattern. The reading of the results was obtained in 24, 48 and 72 h of incubation in a moist chamber, and the definitive reading was in the third period.

3 Results and Discussion

According to the clinical examination, five herds (5 / 9) presented animals malnourished, with respiratory alterations or mastitis, indicating a possible presence of Visna. However, serologic evidences of viral infection were not observed in two hundred ovine serum samples submitted to AGID test for detection of antibodies against Visna in this study (Figure 1).



Figure 1. Negative results in the definitive reading of Agid test after 72 hours of incubation in a moist chamber. PS – positive serum; AG – antigen; SS – serum sample.

A characteristic of SRLV infection is a high prevalence of seropositive animals, apparently healthy, in other words, most of infected small ruminants do not develop clinical signs (PUGH, 2002). In this study, five of nine herds were presenting animals with clinical signs of Visna. However, all the sampled animals were negative by AGID.

Serologic studies for detection of Visna were carried out in the State of São Paulo by Fernandes et al. (2003) that obtained 2.8% of sheep with Visna infection. In the northeast of Brazil, some authors performed serologic studies and the found results were 21.3% and 31.67% of animals seropositive for Visna infection in the States of Rio Grande do Norte (SILVA et al., 2003) and Ceará (ALMEIDA et al., 2003) respectively. However, Oliveira et al. (2006) and Costa et al. (2007) report data in their studies of 5.2% and 1.07% of Visna animal reagents for the state of Pernambuco. Similarly as Araújo et al. (2004) that found 4.93% seropositive sheep for Visna in the State of Ceará. One of the determinants that may explain the different results is the existence of epidemiological or geographical barriers between the areas mentioned in the studies described above and those obtained in the present research. However in the Bahia State, Souza et al. (2007) identified only 0.5% of reagents animals (1 / 200) of two hundred samples of sheep sera from Juazeiro / Bahia, demonstrating the possible presence of Lentivirus in the State of Bahia. And Martinez (2008) also in Juazeiro/BA identified 0.34% of sampled sheep positive by AGID.

Despite these values quantitatively low, the mentionated results above differ greatly from those in this study, since none of all the samples analyzed was reagent for the AGID. Some possible reasons for this result are the lower sensitivity of the Agid assay used, and late seroconversion, and low number of tested animals. Thus, the absence of seropositive animals for Visna in this study is not evidence that this region of the State is free of Visna infection. Besides, there is evidence that cross-reactions are possible for SRLV, indicating a possible presence of Visna in goat herds and the virus spread. Then, further studies are necessary with significant sampling of sheep and goat population, as well as adoption of prevention and control measures to avoid the possible spread of the disease. In this regard, owners of sheep with genetic pattern high and economic value are very important to control the Visna, because the losses would increase with the Visna spread in the Bahia State.

Besides, the difference among the results of this discussion may be justified by a possible modification of the protocol for AGID, as soon as the sensitivity of the method depends on the type of antigen used. According to Adams and Gorham (1986), the antigen produced with glycoprotein 135 (Gp135) has greater sensitivity to the AGID test than the p28 antigen used in this study. Another likely explanation may be due to slow production of antibodies, which can lead to late seroconversion of these animals that may occur months or years after infection; or the animals are at an early stage of infection that the production of antibodies to Visna is absent or low and can not be detected by the assay. This fact raises the frequency of false-negative results for Visna (LEROUX et al., 1997; FLEET et al., 2005; ELTAHIR et al., 2006), being a major problem in animals trade, because may favor the spread of the disease unnoticed inter-herds.

Furthermore, the AGID test and the enzymelinked immunosorbent assay (Elisa), both are prescribed tests for international trade by the World Organisation for Animal Health (OIE, 2004). However the AGID is reported to be more specific, reproducible and simple to perform, but requires training and experience in reading the results (DE ANDRES et al., 2005). The Elisa is sensitive, cheaper and stages of the process can be automated (SAMAN et al., 1999), therefore making it useful for screening large numbers of sera. Thus, none of the two tests are perfects; the optimal choice of test depends on the purpose of testing, whether the objective is surveillance of a population or confirmation of suspected cases. Thereby, Toft et al. (2007) combined the AGID and commercially available indirect Elisas tests in a serial or parallel reading to improve the performance of diagnostic tests and found no significant improvement in the specificity, but the sensitivity was reduced significantly.

It is important to emphasize the hypothesis on the presence or absence of Visna in herds of the Recôncavo Baiano, because despite the absence of seropositive sheep in this research, recent phylogenetic studies have demonstrated the transmission of SRLV of sheep to goats and vice versa (LEROUX et al., 1997; SHAH et al., 2004; GJERSET et al., 2009; GLARIA et al., 2009). Based on these studies is not daring to speculate that the MVV can be present in small ruminants herds in the State of Bahia.

4 Conclusions

Therefore, one concludes that the animals sampled had no evidence of antibodies to Visna IDAG detectable by using viral p28 antigen at the collection period. Nevertheless, to confirm the absence of *Visna/Maedi* in sheep herds in Bahia, there is a need for further studies with significant sampling including a more sensitive assay.

Acknowledgments

We thanks Fapesb (Fundação de Amparo á Pesquisa do Estado da Bahia) for the research grant, and Pibic-Ufba (Programa Institucional de Bolsas de Iniciação Científica da Universidade Federal da Bahia) for the scientific initiation. This article is dedicated to Professor Eugenia Márcia de Deus Oliveira (In memoriam).

References

ADAMS, D.S.; GORHAM, J.R. The gp135 of caprine arthritis encephalitis virus affords greater sensitivity than the p28 in immunodiffusion serology. *Research in Veterinary Science*, v.40, p.157-160, 1986.

ALMEIDA, N.C.; TEXEIRA, M.F.S.; FERREIRA, R.C.S.; CALLADO, A.K.C.; FROTA, M.N.; MELO, A.C.M.; APRIGIO, C.J.L. Detecção de ovinos soropositivos para Maedi/Visna destinados ao abate na região metropolitana de Fortaleza. *Veterinária Notícias*, v.9, n.1, p.59-63, 2003.

ARAÚJO, S.A.C.; DANTAS, T.V.M.; SILVA, J.B.A.; RI-BEIRO, A.L.; RICARTE, A.R.F.; TEIXEIRA, M.F.S. Identificação do Maedi-Visna vírus em pulmão de ovinos infectados naturalmente. *Arquivo do Instituto Biológico*, São Paulo, v.71, n.4, p.431-436, out./dez., 2004.

BLACKLAWS, B.A.; BERRIATUA, E.; TORSTEINSDOTTIR, S.; WATT, N.J.; De ANDRÉS, D.; KLEIN, D.; HARKISS, G.D. Transmission of small ruminant lentiviruses. *Veterinary Microbiology*, v.101, n.3, p.199-208, 2004.

CONCHA-BERMEJILLO, A. de La; MAGNUS-CORRAL, S.; BRODIE, S.J.; DeMARTINI, J.C. Venereal shedding of ovine lentiviruss in infected rams. *American Journal of Veterinary Research*, v.57, p. 684-688, 1996.

COSTA, L.S.P.; LIMA, P.P. de; CALLADO, A.K.C.; NAS-CIMENTO, S.A.do; CASTRO, R.S. Lentivírus de pequenos ruminantes em ovinos Santa Inês: isolamento, identificação pela PCR e inquérito sorológico no Estado de Pernambuco. *Arquivo do Instituto Biológico*, São Paulo, v.74, n.1, p.11-16, jan./mar., 2007.

DE ANDRES, D.; KLEIN, D.; WATT, N.J.; BERRIATUA, E.; TORSTEINSDOTTIR, S.; BLACKLAWS, B.A.;

- HARKISS, G.D. Diagnostic tests for small ruminant lentiviruses. *Veterinary Microbiology*, v.107, p.49–62, 2005.
- ELTAHIR, Y.M.; DOVAS, C.I.; PAPANASTASSOPOU-LOU, M.; KOUMBATI, M.; GIADINIS, N.; VERGHESE-NIKOLAKALI, S.; KOPTOPOULOS, G. Development of a semi-nested PCR using degenerate primers for the generic detection of small ruminant lentivirus proviral DNA. *Journal of Virological Methods*, v. 135, p. 240-246, 2006.
- FERNANDES, M.A.; ARAUJO, W.P.; CASTRO, R.S. Prevalência da infecção pelo vírus Maedi-Visna em ovinos da microrregião da grande São Paulo, Estado de São Paulo. *Ciência Veterinária nos Trópicos*, v.6, n.1, p.23-28, 2003.
- FROTA, M.N.L.; SILVA, J.B.A.; ARAUJO, S.A.C.; TEIXEIRA, M.F.S. Artrite Encefalite Caprina em cabritos de rebanhos com programa de controle no Estado do Ceará. *Arquivo do Instituto Biológico*, São Paulo, v.72, n.2, p.147-152, abr./jun.,2005.
- GJERSET, B.; RIMSTAD, E.; TEIGE, J.; SOETAERT, K.; JONASSEN, C.C. Impact of natural sheep-goat transmission on detection and control of small ruminant lentivirus group C infections. *Veterinary Microbiology*, v.135, p.231-238, 2009.
- GLARIA, I.; REINA, R.; CRESPO, H.; de ANDRÉS, X.; RAMÍREZ, H.; BIESCAS, E; PÉREZ, M.M.; BADIOLA, J.; LUJÁN, L.; AMORENA, B.; de ANDRÉS, D. Phylogenetic analysis of SRIV sequences from an arthritic sheep outbreak demonstrates the introduction of CAEV-like viruses among Spanish sheep. Veterinary Microobiology, v.138, p.156-162, 2009.
- LEGINAGOIKOA, I.; DALTABUILT-TEST, M.; ÁLVAREZ, V.; ARRANZ, J.; JUSTE, R.A.; AMORENA, B.; DE ANDRÉS, D.; LUJÁN, J.J.; BADIOLA, J.J.; BERRIATUA, E. Horizontal Maedi-Visna virus (MVV) infection in adult dairy- sheep raised under varying MVV-infection pressures investigated by ELISA and PCR. Research in Veterinary Science, v. 80, p. 235-241, 2006.
- LEROUX, C.; LERONDELLE, C.; CHASTANG, J.; MORNEX, J.F. RT-PCR detection of lentiviruses in milk or mammary secretion of sheep or goats from infected flocks, *Veterinary Research*, v.28, p.115-121, 1997.
- MARTINEZ, P.M.; Características dos sistemas de produção de ovinos e prevalência sorológica da Maedi-Visna na Microrregião de Juazeiro-Bahia. Bahia, Brasil. 82f., 2008. Dissertação (Mestrado) Universidade Federal da Bahia. Salvador, 2008.
- MELO, C.B.; CASTRO,R.S.; OLIVEIRA, A.A.; FONTES, L.B.; CALLADO, A.K.; NASCIMENTO, S.A.; MELO, L.E.H.; SILVA, J.S. Estudo preliminar sobre a infecção por lentivírus de pequenos ruminantes em ovinos e caprinos em Sergipe. In: CONGRESSO LATINOAMERICANO, 11; CONGRESSO BRASILEIRO, 5; CONGRESSO NORDESTINO DE BUIATRIA, 3. Salvador, 2003. Anais. p.47.

- MOOJEN, V.; BARTH, O. M.; RAVAZZOLO, A.P.; GROLL, ANDREA Von; CORTES, L.M.C.; MARCHESIN, D.M. Sheep Maedi-Visna Virus: Ultrastructural Characterization of A Brazilian Isolate. In: VIROLÓGICA 95, 1995, Ribeirão Preto. Anais. Ribeirão Preto SP, 1995. p. B14.
- OIE. World Oraganization for Animal Health. Manual of diagnostic tests and vaccines for terrestrial animals. 2004. 5ed. Available at: http://www.oie.int. Accessed 30 april 2007
- PASICK, J. Maedi-Visna virus and Caprine Arthritis-Encephalitis Virus: distinct species or quasispecies and its implications for laboratory diagnosis. *Canadian Journal* of Veterinary Research, Montreal, v.62, p.241-244, 1998.
- PREZIUSO, S.; SANNA, E.; SANNA, M.P.; LODDO, C.; CERRI, D.; TACCINI, E.; MARIOTTI, F.; BRAGA, G.; ROSSI, G.; RENZONI, G.; Association of *Maedi Visna* virus with *Brucella ovis* infection in rams. *European Journal of Histochemistry*, v.47, p.151-157, 2003.
- PUGH, D.G. Ovine Progressive Pneumonia. In: Sheep and Goat Medicine. W.B. Saunders Company. p. 239-141, 2002.
- RAVAZZOLO, A.P.; MARCHESIN, D.M.; CALDAS, A.P.F.; VIEIRA, L.A.; MOOJEN, V.; QUÉRAT, G. Detection of Brazilian Isolates of Visna-Maedi And Caprine Arthritis-Encephalitis Virus by Polymerase Chain Reaction. In: VIROLÓGICA 95, 1995, Ribeirão Preto. *Anais*, 1995. p. B13.
- RAVAZZOLO, A.P.; REISCHAK, D.; PETERHANS, E. Zanoni, R. Phylogenetic analysis of small ruminant lentiviruses from Southern Brazil. *Virus Research*, v. 79, p. 117-123, 2001.
- REINA, R.; BERRITUA, E.; LUJÁN, L.; JUSTE, R.; SÁNCHEZ, A.; de ANDRÉS, D.; AMORENA, B. Prevention strategies against small ruminant lentiviruses: an update. *The Veterinary Journal*, v. 182, p.31-37, 2009.
- SAMAN, E.; VAN EYNDE, G.; LUJAN, L.; EXTRAMIANA, B.; HARKISS, G.; TOLARI, F.; GONZALEZ, L.; AMORENA, B.; WATT, N.; BADIOLA, J. A new sensitive assay for detection of lentivirus infections in small ruminants. *Clinical and Diagnostic Laboratory Immunology* v.6, p.734–740, 1999.
- SHAH, C.; BONI, J.;HUDER, J.B.; VOGT, H.-R.; MUHLHERR, J.; ZANONI, R.; MISEREZ, R.; LUTZ, H.; SCUPBACH, J. Phylogenetic analysis and reclassification of caprine and ovine lentiviruses based on 104 new isolates: evidence for regular sheep-to-goat transmission and worldwide propagation though livestock trade. *Virology*, v.319, p.12-26, 2004.
- SILVA, J.B.A. Levantamento sorológico pelo teste de imunodifusão em gel de agarose (IDGA) da lentivirose ovina em rebanhos do Rio Grande do Norte, Brasil, 2003. 58f. Dissertação (Mestrado) Faculdade de Veterinária, Univ. Estadual do Ceará, Fortaleza, 2003.

SILVA, J.S.; CASTRO, R.S.; MELO, C.B.; FEIJÓ, F.M.C. Infecção pelo vírus da artrite encefalite caprina no Rio Grande do Norte. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, v.57, n.6, p.726-731, 2005.

SMITH, M.C.; SHERMAN, D.M. Maedi-Visna (MV) and Caprine Arthritis Encephalitis. In: Goat Medicine. p. 73-79; 135-138, 1994.

SOUZA, T.S. de; COSTA, J.N.; MARTINEZ, P.M.; PINHEIRO, R.R. Estudo sorológico da Maedi-Visna pelo método da Imunodifusão em Gel de ágar em rebanhos ovinos de Juazeiro, Bahia, Brasil. Revista Brasileira de Saúde e Produção Animal, v.8, n.4, p.276-282, out./dez. 2007.

TOFT, N.; AKERSTEDT, J.; THARALDSEN, J.; HOPP, P. Evaluation of three serological tests for diagnosis of Maedi-Visna virus infection using latent class analysis. *Veterinary Microbiology*, v.120, p.77-86, 2007.